

University of Groningen

Cross-validation of Liquid Chromatography-Tandem Mass Spectrometry Method for Quantification of Levofloxacin in Saliva

Ghimire, Samiksha; Jongedijk, Erwin M.; van den Elsen, Simone H.J.; Wessels, Mireille A.; Touw, Daan J.; Alffenaar, Jan-Willem C.

Published in:
Journal of Applied Bioanalysis

DOI:
[10.17145/jab.20.008](https://doi.org/10.17145/jab.20.008)

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version
Publisher's PDF, also known as Version of record

Publication date:
2020

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

Ghimire, S., Jongedijk, E. M., van den Elsen, S. H. J., Wessels, M. A., Touw, D. J., & Alffenaar, J-W. C. (2020). Cross-validation of Liquid Chromatography-Tandem Mass Spectrometry Method for Quantification of Levofloxacin in Saliva. *Journal of Applied Bioanalysis*, 6(2), 68-70. <https://doi.org/10.17145/jab.20.008>

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

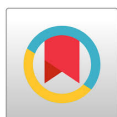
Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Downloaded from the University of Groningen/UMCG research database (Pure): <http://www.rug.nl/research/portal>. For technical reasons the number of authors shown on this cover page is limited to 10 maximum.

LETTER TO EDITOR

Cross-validation of Liquid Chromatography-Tandem Mass Spectrometry Method for Quantification of Levofloxacin in Saliva



Citation:

Ghimire S, Jongedijk EM, van den Elsen SH, Wessels MA, Touw DJ, Alffenaar JW. Cross-validation of Liquid Chromatography-Tandem Mass Spectrometry method for quantification of Levofloxacin in saliva. *Appl Bioanal* 6(2), 68-70 (2020).

Editor:

Dr. Roland J.W. Meesters,
MLM Medical Labs GmbH,
Moenchengladbach, Germany.

Received: February 17, 2020.

Revised: May 27, 2020.

Accepted: June 02, 2020.

Open Access & Copyright:

©2020 Ghimire S et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/) (CC-BY) which permits any use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

Funding & Manuscript writing assistance:

The authors have no financial support or funding to report and they declare that no writing assistance was utilized in the production of this article.

Financial & Competing interests:

The authors have declared that no competing interest exist.

Samiksha Ghimire^{1,*}, Erwin M. Jongedijk¹, Simone H.J. van den Elsen¹, Mireille A. Wessels¹, Daan J. Touw^{1,2}, Jan-Willem C. Alffenaar^{1,3,4}

¹University of Groningen, University Medical Center Groningen, Department of Clinical Pharmacy and Pharmacology, Groningen, The Netherlands; ²University of Groningen, Groningen Research Institute of Pharmacy, Department of Pharmacokinetics Toxicology and Targeting, Groningen, The Netherlands; ³University of Sydney, Faculty of Medicine and Health, School of Pharmacy, Sydney, Australia; ⁴Westmead hospital, Sydney, Australia.

***Correspondence:** Samiksha Ghimire, PhD, University of Groningen, University Medical Center Groningen, Clinical Pharmacy and Pharmacology, Groningen, The Netherlands. Hanzeplein 1, 9713 GZ Groningen; Phone: +31 (0)615 547 345; Email: samixa7@gmail.com

Keywords: levofloxacin, saliva, liquid chromatography tandem mass spectrometry, tuberculosis.

Levofloxacin belongs to the Group A drug for treating multi-drug resistant tuberculosis (MDR-TB) but exhibits considerable pharmacokinetic variability. For a 750-1000 mg once daily dosing, the desired levofloxacin plasma/serum concentration range is 8-12 mg/L and the area under the concentration time curve from 0 to 24h is 75 if MIC is 0.5 mg/L and 150 if MIC is 1 mg/L. Saliva too could be a potential patient friendly alternative sampling matrix for levofloxacin quantification [1,2]. However, levofloxacin quantification in saliva using a liquid chromatography-tandem mass spectrometry (LC-MS/MS) method developed for plasma or serum requires cross validation. Moreover, the handling of infectious saliva samples from TB patients puts health care workers at risk of contagion. Membrane filtration was found to be suitable for sterilization of saliva samples [3]. The aims of this study were: a) to assess if drug concentrations in human saliva could be reliably determined with calibration samples prepared in human serum; and b) to perform a recovery test for levofloxacin concentrations in saliva after using sorbent material such as cotton rolls and/or filtering through a membrane filter.

A slight modification was done to our previously published LC-MS/MS method for levofloxacin quantification in human serum/plasma [4]. The assay was adjusted to simultaneously detect ciprofloxacin, moxifloxacin and levofloxacin in plasma/serum. First, for cross validation, levofloxacin stock solution of 2.5 mg/mL was prepared in dimethyl sulfoxide (Merck, NJ, USA). Nine different concentrations of the calibration samples in blank human serum were made: 0.20, 0.50, 1, 2, 5, 10, 20, 40, and 50 mg/L. In addition, four different concentrations of quality control samples (QC) in saliva, with a lower limit of quantification at 0.2 mg/L, low QC at 1 mg/L, medium at 20 mg/L, and high at 40 mg/L were prepared. The internal standard solution was prepared from a 1 mg/ml stock solution of [³H₄]-levofloxacin in DMSO by diluting 50 µl to 250 ml with methanol (0.2 mg/L). For cross validation, all samples were analyzed in quintuplicate. The analysis was performed on a triple quadrupole LC-MS/MS (Thermo Scientific TSQ Quantiva, San Jose, CA, USA). A Thermo Accucore C18 analytical column of particle size 2.6 µm, 50 mm length, and internal diameter of

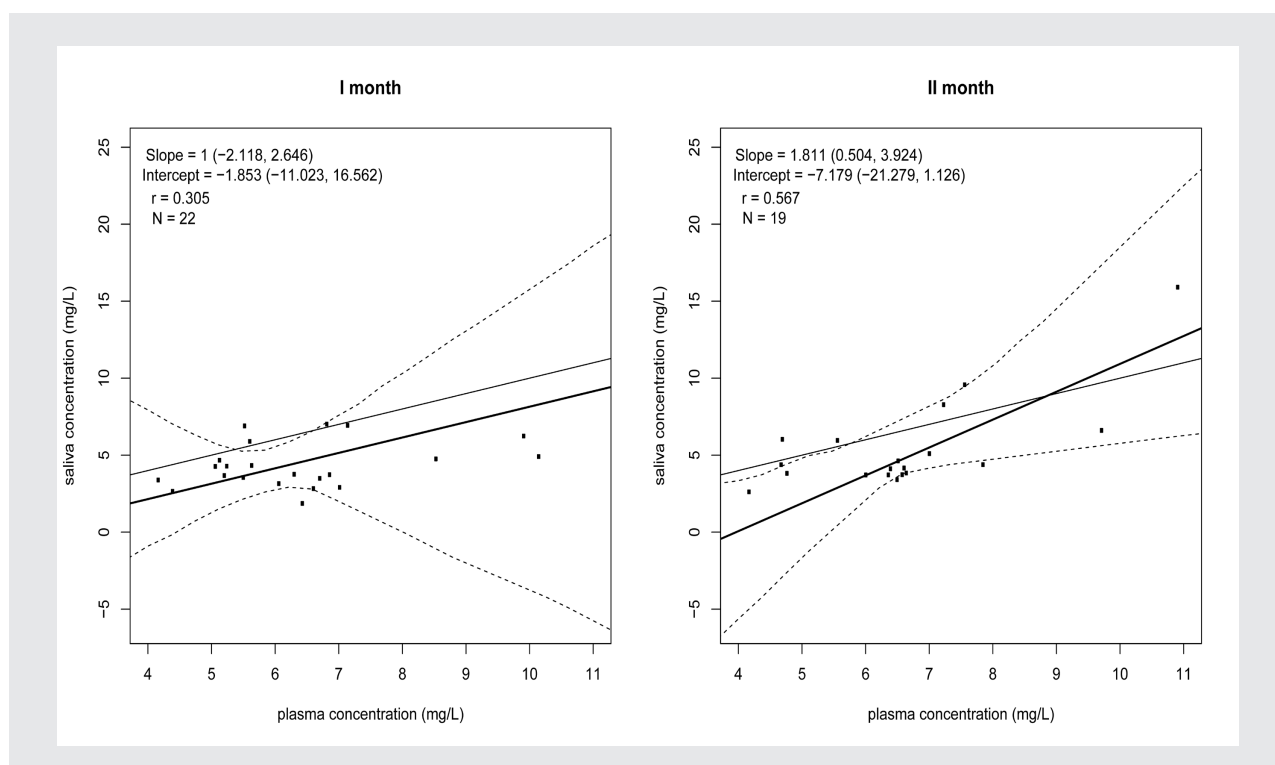


Figure 1. Passing-Bablok regression analysis of mean Lfx concentrations (at 0, 1, 2, 4, and 8 h) in plasma and saliva for 2 months. The bold solid line represents the Passing-Bablok fitted line, whereas the solid lighter line is the line of identity. The dashed lines indicate the 95% CI, r is the Spearman's rank correlation, and N is the number of paired mean plasma and saliva concentrations. Reprinted with permission from [5]. ©American Society for Microbiology (2019).

2.1 mm was used. The column temperature during analysis was 60°C. The linearity of the calibration curve was 0.20–50 mg/L for levofloxacin in both serum and saliva. QC samples in saliva at four concentration levels (0.20, 1, 20, 40 mg/L) were quantified using a calibration curve in serum. All QC samples were prepared and measured in 5-fold during a single day. The LC-MS/MS method had a run time of 2 min and levofloxacin eluted at a retention time of 0.7 min. Accepted bias and coefficient of variation (CV) were $\leq 15\%$ for QC samples at low (at -0.9% and 1.0%), medium (at -0.3% and 0.9%), and high (at 2.0% and 1.3%) concentrations and $\leq 20\%$ for LLOQ (at -1.0% and 2.3%) in saliva. This method was clinically applied for the analysis of levofloxacin concentrations in saliva samples at the laboratory of the department of Clinical Pharmacy and Pharmacology in the University Medical Center Groningen for a clinical trial (identifier number [NCT 03000517](#)) on the pharmacokinetics of levofloxacin in saliva of 23 MDR-TB patients. The median observed AUC_{0-24} and C_{max} in saliva were 67.09 mg·h/L and 7.03 mg/L [5]. Levofloxacin concentrations in plasma and saliva of 23 MDR-TB patients is shown (Figure 1).

Second, the recovery of levofloxacin in saliva was evaluated using four different solutions. The first group (blank syringe), was blank saliva which was absorbed by the cotton roll and afterwards compressed in a syringe. The effluent was then spiked with levofloxacin at 1 and 5 mg/L. In the second group (test solution syringe), levofloxacin spiked saliva at concentrations of 1 mg/L and 5 mg/L were applied to the cotton rolls. The volume required to saturate the cotton rolls was determined beforehand. Thereafter, cotton rolls with absorbed spiked saliva were compressed in a syringe by pushing the plunger of the syringe and collecting the effluent. The recovery was evaluated in the effluent using the

blank syringe solution as a reference. The third group (blank syringe filter) was similar to the first group, except the blank saliva was pushed through the syringe equipped with a 0.22 μm polyvinylidene fluoride membrane filter, and later spiked with levofloxacin at the above-mentioned concentrations. In the fourth group (test solution syringe filter), recovery yield was determined after compressing fully saturated cotton rolls with levofloxacin spiked saliva at (1 mg/L and 5 mg/L) in a syringe equipped with a 0.22 μm membrane filter. The blank syringe filter solution was used as a reference to determine the recovery. Our study has shown that the plain cotton rolls achieved a recovery of around 70% at 1 mg/L with a CV% of 9.5%; whereas at 5 mg/L the mean recovery was more variable between the groups (63-80%) with a CV of 6.0%. This will have an impact on the variability of analytical results with a spread of 17% and bias of approximately 30%, if cotton rolls are used as a sampling device. This is likely due to sorption of levofloxacin to the cotton roll. Therefore, saliva samples could be useful only in screening and semi-quantitative prediction of plasma levels of anti-TB drugs [5]. In addition, our experiments have shown that filtration through a 0.22 μm polyvinylidene fluoride membrane filter does not result in a further loss of levofloxacin.

In conclusion, results of cross-validation study were within the acceptance criteria for bias and precision according to formal regulations. The cotton rolls used for saliva sample collection achieved a levofloxacin recovery of around 70%.

REFERENCES

1. Gröschl M. Saliva: a reliable sample matrix in bioanalytics. *Bioanalysis* 9, 655-668 (2017).
2. van den Elsen, Simone HJ, Oostenbrink LM, Heysell SK, et al. Systematic Review of Salivary Versus Blood Concentrations of Antituberculosis Drugs and Their Potential for Salivary Therapeutic Drug Monitoring. *Ther Drug Monit* 40, 17-37 (2018).
3. van den Elsen SHJ, van der Laan T, Akkerman OW, et al. Membrane Filtration Is Suitable for Reliable Elimination of Mycobacterium tuberculosis from Saliva for Therapeutic Drug Monitoring. *J Clin Microbiol* 55, 3292-3293 (2017).
4. Ghimire S, van Hateren K, Vrubleuskaya N, et al. Determination of levofloxacin in human serum using liquid chromatography tandem mass spectrometry. *J Appl Bioanal* 4(1), 16-25 (2018).
5. Ghimire S, Maharjan B, Jongedijk EM et al. Evaluation of Saliva as a Potential Alternative Sampling Matrix for Therapeutic Drug Monitoring of Levofloxacin in Patients with Multidrug-Resistant Tuberculosis. *Antimicrob Agents Chemother* 23 (5) e02379-18 (2019).